

# Data integration of highly dimensional biological data sets with multivariate analysis

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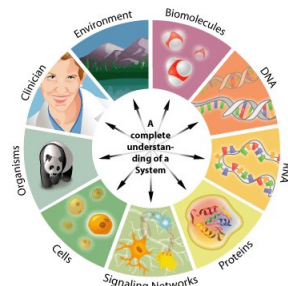
Queensland Facility for Advanced Bioinformatics  
The University of Queensland



- Study of complex interactions in biological systems
- Holism vs. reductionism
 

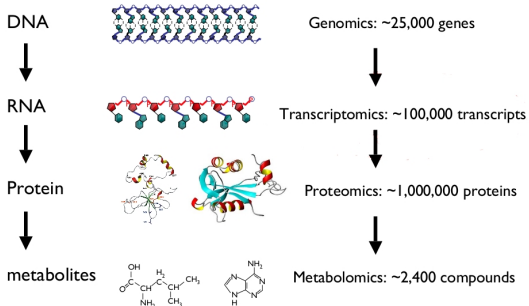
*'Systems biology [...] requires that we develop ways of thinking about integration that are as rigorous as our reductionist programmes, but different [...].It means changing our philosophy, in the full sense of the term',*

Denis Noble
- **Biology-based inter-disciplinary study field**



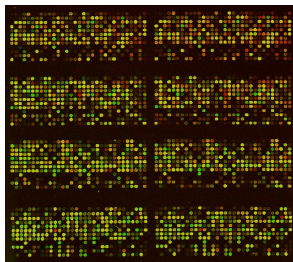
- Understand better the entirety of processes that happen in a biological system
- Model and discover emergent properties, properties of cells, tissues and organisms functioning as a system

# The biological dogma and the 'omics' cascade



→ **Integrative systems biology**: understand the relationships between these functional levels

# Transcriptomics: DNA microarray technology



- Measures the expression of thousands of genes on a single individual
- 1 spot = 1 gene
- Gene expression measure = signal intensity
- Spots are 'on' (activated) or 'off' (silent) across biological conditions

→ Identify biomarkers or regulated genes to understand the processes of cellular differentiation or carcinogenesis  
(Supervised analysis)

# High-throughput sequencing

**DNA sequencing:** methods for determining the order of the nucleotide bases A-C-G-T in a molecule of DNA.

**High-throughput sequencing:** parallelize the sequencing process, produces thousands or millions of sequences at once

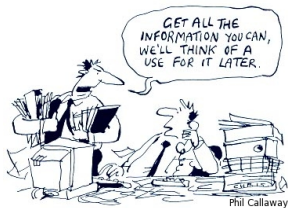


- inexpensive genome-wide sequence and fast
- provides insights into genome variation and evolution
  - genotyping, genome resequencing, de novo genome assembly projects and metagenomics
  - need of efficient methodologies to process and analyse the data



# Challenges

## Close interaction between statisticians, bioinformaticians and molecular biologists



- Understand the biological problem
- Irrelevant (noisy) variables
- $n \ll p$  and  $n$  very small  
→ limited statistical validation
- Is the statistical approach is biologically relevant?
- Keep up with new technologies
- Anticipate computational issues

# Some research questions

Now, consider the framework of **longitudinal 'omics' studies** ...

- 1 Do we observe a **'natural' separation** between the different groups of patients **across time**?
- 2 **Cluster the times profiles** for:
  - same type** of biological features
  - different type** of biological features
  - **Identify subsets of correlated features across time**
- 3 Do **several assays** performed on the same samples contain the **same information**?

## Linear Multivariate approaches

- Dimension reduction  
→ [project](#) the data in a smaller subspace
- To handle multicollinear, irrelevant, missing variables
- To capture experimental and biological variation

In the R package [mixOmics](#), focus is on:

- Data integration
- Variable selection
- Computationally efficient methodologies for large biological data sets
- Interpretable graphical outputs



# Principal Component Analysis: PCA

Seek the best directions in the data that account for most of the variability

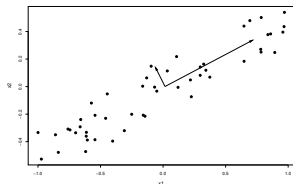
→ **principal components**: artificial variables that are linear combinations of the original variables:

$$\mathbf{c}_1 = w_1\mathbf{x}_1 + w_2\mathbf{x}_2 + \dots + w_p\mathbf{x}_p$$

where

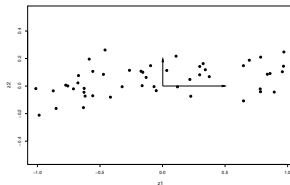
- where  $\mathbf{c}_1$  is the **first** principal component with max. variance
- $\{w_1, \dots, w_p\}$  are the weights in the linear combination
- $\{\mathbf{x}_1, \dots, \mathbf{x}_p\}$  are the gene expression profiles.

All PCs are mutually orthogonal. ( $\mathbf{c}_1, \mathbf{c}_2, \dots$ )



The new PCs form a smaller subspace of dimension  $< p$

Project the data on these new axes to summarize the information related to the variance.



→ approximate representation of the data points in a lower dimensional space

PCA is an (almost) compulsory first step in exploratory data analysis to:

- Have a first understanding of the underlying data structure
- Identify bias, experimental errors, batch effects

Problem with PCA: interpretation can be difficult with very large number of (possibly) irrelevant variables.

Remember that the principal components are linear combinations of the original variables:

$$c = w_1x_1 + w_2x_2 + \dots + w_px_p$$

A clearer signal could be observed if some of the variable weights  $\{w_1, \dots, w_p\}$  could be set to 0 for the irrelevant variables:

$$c = 0 * x_1 + w_2x_2 + \dots + 0 * x_p$$

These variables weights are defined in the loading vectors.  
Important weights = important contribution to the PC.  
Similar weights = correlated variables.

# sparse Principal Component Analysis: sPCA

sparse PCA: **sparse loading vectors** to remove noisy or irrelevant variables which determine the principal components.

→ Solving PCA through least squares problem (SVD) allows to include regularization parameters

$$\min_{\mathbf{v}_h} \|\mathbf{X}_h - \mathbf{u}_h \mathbf{v}_h^T\|_F^2 + P_\lambda(\mathbf{u}_h)$$

$P_\lambda$  is a penalty function with tuning regularization parameter  $\lambda$

→ use **Lasso** penalization, or soft-thresholding

→ obtain **sparse loading vectors**, with very few non-zero elements

**Shen, H., Huang, J.Z.** 2008. **Sparse principal component analysis via regularized low rank matrix approximation**, *J. Multivariate Analysis*.

## Why PCA can 'fail' to summarize the data?

- In some time course experiments, the **subject variation can be larger than the time variation**
- PCA makes the assumption that samples are **independent** of each other
- In univariate analysis we use a paired t-test instead of a t-test
- In multivariate analysis we use a **multilevel** approach:
  - different sources of variation can be separated (**treatment effect within subjects** and differences between subjects)
  - gain in power

## Multilevel approach

- The variation in the data is separated: within matrix and between matrix
- Multivariate tools can then be applied on the within matrix (Westerhuis, 2008)
- We can take into account the **repeated measures design** of the experiment

**VEGFC Study:** Human lymphatic endothelial cells were treated in vitro with recombinant VEGF-C for **16 time points**: 0min, 15min, 30min, 45min, 60min, 80min, 100min, 2h, 2.5h, 3h, 3.5h, 4h, 5h, 6h, 7h, or 8h) in **triplicates**, **CAGE data** (FANTOM5, Riken Institute).

**Liquet\*, B., Lê Cao\*, K-A., Hocini, H., Thiébaud, R.** **A novel approach for biomarker selection and the integration of repeated measures experiments from two platforms**, *BMC Bioinformatics*, accepted (25/09/2012).

# VEGFC study: high individual effect

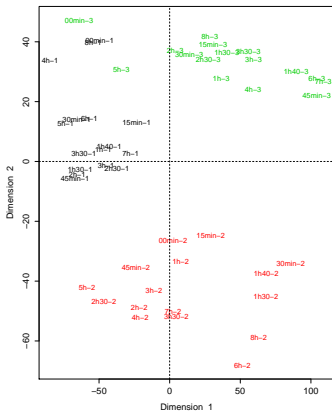


Figure: PCA on original data, color = patient

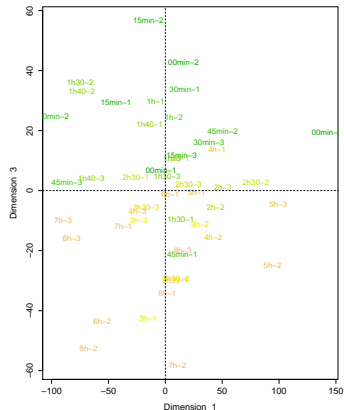


Figure: PCA within matrix, color = time

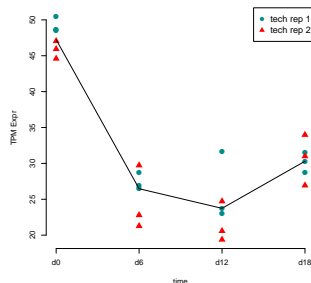


## Modelling trajectories: cubic smoothing splines

Aim: summarize the trajectory of each variable

- Use cubic smoothing splines to summarize each profile
- The **derivative** between each time point can be estimated
- Fit a non-supervised algorithm to cluster the profiles based on the derivative (k-means, SOM)

Example on one CAGE cluster



Déjean et al. (2008), Clustering Time-Series Gene Expression Data Using Smoothing Spline Derivatives *Eurasip J.*



# Example with K-means

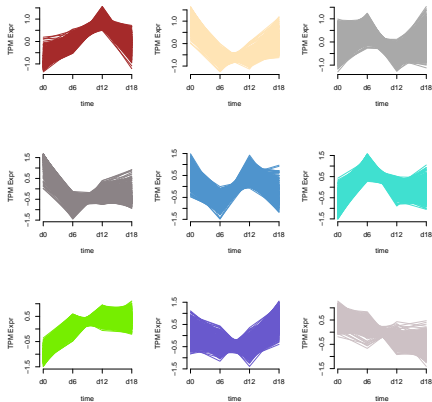


Figure: K-means on derivatives

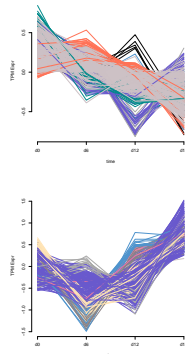


Figure: K-means on original data

# How about with Self Organising Maps (SOM)?

Time profiles

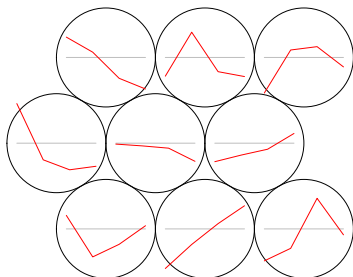


Figure: SOM summary

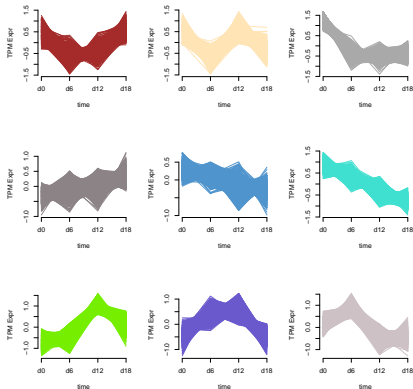


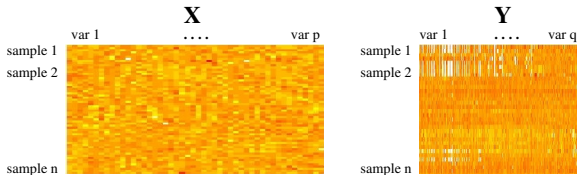
Figure: SOM on original data

## Link between smoothing splines and LMM

- Be able to assess the variability for each feature:
  - technical variability
  - biological variability
- Well fitted for correlated repeated measures
- Fits into a **linear mixed model framework** (not parameters to tune, Verbyla et al. 1999)
- Can take into account random intercepts, bio reps as random effects ...
- Enables interpolation of **missing values**
- Enables to **model the shape of the trajectories**
- Variance components and estimates of fixed and random effects can be obtained

## Integrating two large data sets

Aim: **integrate** two data sets and **select** the relevant features simultaneously:



- Two large data sets X and Y
- Measurements of **two types** of *variables* on the **same samples**

- Partial Least Squares regression **maximises the covariance** between each linear combination (components) associated to each data set

$$\max_{\|u_h\|=1, \|v_h\|=1} \text{cov}(X_h u_h, Y_h v_h), \quad h = 1 \dots H$$

where  $X$  ( $n \times p$ ) is the transcriptomics data set and  $Y$  ( $n \times q$ ) is the proteomics data set

- Similarly to PCA, the **PLS components** indicate the **similarities between samples** (useful plots!)
- The **loading vectors** indicate the **contribution** of the variables of the same type to the PLS component (useful for variable selection)

## sparse PLS-SVD

Use the PLS-SVD variant that directly gives the latent variables and loading vectors and low rank rank approximation.

Let  $M_h = X_h^T Y_h$ , **sparse PLS** solves the optimization problem:

$$\min_{\mathbf{u}_h, \mathbf{v}_h} \|M_h - \mathbf{u}_h \mathbf{v}_h'\|_F^2 + P_{\lambda_1}(\mathbf{u}_h) + P_{\lambda_2}(\mathbf{v}_h)$$

where  $P_\lambda$  is a penalty function

- obtain simultaneously **sparse** loadings  $\mathbf{u}_h$  and  $\mathbf{v}_h$
- **simultaneous select variables from both data sets which are correlated across samples**

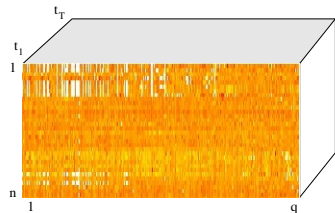
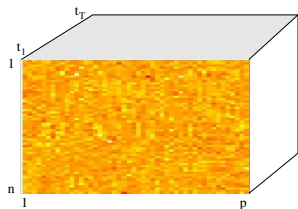
**Lê Cao K-A., Rossouw D., Robert-Granié C. and Besse P.** 2008. **A Sparse PLS for Variable Selection when Integrating Omics data.** *SAGMB* **7**(1).

## Parameters to tune

- Number of PLS components:
  - $Q_h^2$  index
  - graphical outputs
- Lasso penalizations  $\lambda_1^h, \lambda_2^h$  ( $h = 1, \dots, H$ ):
  - error prediction with cross-validation
  - maximisation of the covariance, stability analysis, permutations(?)

→ the biologist will also help choosing these parameters!

## Integration of two longitudinal studies



- Select correlated profiles across time, between and within each data set.
- But difficult to deal with 3D data sets!
- PLS can integrate 2 data sets of 2 dimensions.



Step 1: use cubic smoothing splines to reduce one dimension (samples dimension)

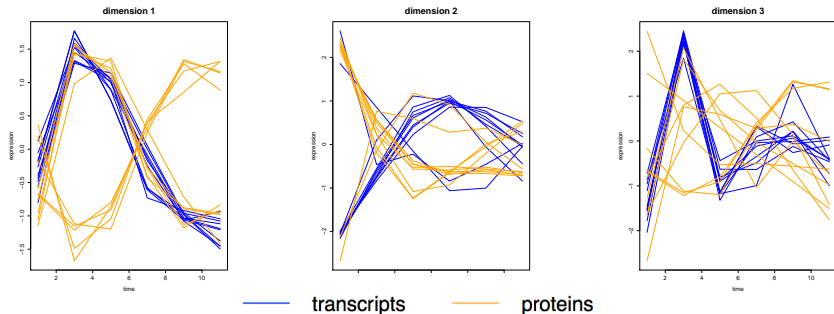
Step 2: apply sPLS on the estimated splines to identify correlated profiles both within and between the two data sets

Two illustrative studies:

**Kidney transplant study:** Transcriptomics and proteomics study of 40 patients with kidney transplant, rejecting ( $n_1 = 20$ ) or not ( $n_2 = 20$ ) the transplant. Follow up on 5 time points (weeks), PROOF Centre, UBC.

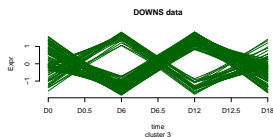
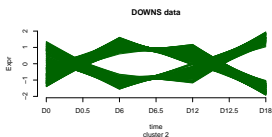
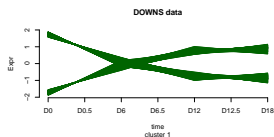
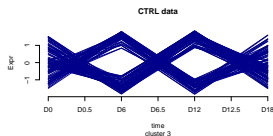
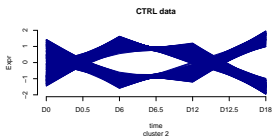
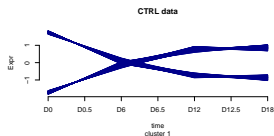
**Neuronal study:** Human induced pluripotent stem cells from Downs syndrome patients and controls differentiated to neurons (CAGE data). 2 bio reps and 3 tech reps per genotype (control, down syndrome), 4 time points (days), FANTOM5, Riken Institute .

## Profile clusters on kidney transplant study



- sPLS selects both transcripts and proteins which are **positively or negatively correlated** across time
- Quality of clusters decreases with the number of PLS components (dimensions) as **obvious patterns cannot be extracted anymore**

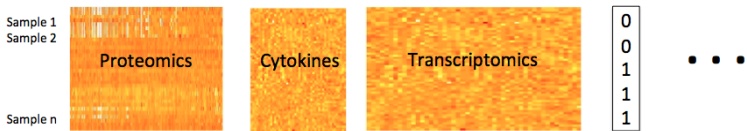
## Concordant profile clusters on Neuronal study



- Each cluster of profiles corresponds to a PLS component.
- Selection of **different** features per condition and per component.

# Integration of multiple data sets

Integrate heterogeneous data sets



- Need to define the relationships between the different data sets
- **Select relevant biological entities** which are correlated across the different data sets

## Regularized CCA

Classical Canonical Correlation Analysis solves the problem

$$\max \text{cor}_{\mathbf{a}_h, \mathbf{b}_h}(\mathbf{X}\mathbf{a}_h, \mathbf{Y}\mathbf{b}_h) \quad \text{s.t.} \quad \text{var}(\mathbf{X}\mathbf{a}_h) = \text{var}(\mathbf{Y}\mathbf{b}_h) = 1$$

For  $n \ll p + q$ , the empirical covariance matrices are **ill-conditioned**  $\rightarrow$  canonical correlations close to 1.

In **regularized CCA** the covariance matrices are replaced by:

$$\text{Cov}(\mathbf{X}) + \lambda_1 \mathbf{Id} \quad \text{and} \quad \text{Cov}(\mathbf{Y}) + \lambda_2 \mathbf{Id}$$

**González I., Déjean S., Martin P.G.P., Goncalves O., Besse P. and Baccini A. 2009** Highlighting Relationships Between Heterogeneous Biological Data Through Graphical Displays Based On Regularized Canonical Correlation Analysis, *Journal of Biological Systems*, 17 (2).

## Multi-block analysis: Regularized Generalised CCA

- RGCCA generalizes rCCA to **more than 2 data sets**
- Constitutes a **general framework** for many multi-block data analysis methods
- Objective: seeks linear combinations of block variables:
  - (i) block components explain their own block well and/or
  - (ii) block components that are assumed to be connected are highly correlated.

**Tenenhaus, A., Tenenhaus, M (2011)** [Regularized Generalised Canonical Correlation Analysis](#), *Psychometrika*, 76 (2).

# RGCCA

For  $J$  blocks of variables  $\mathbf{X}_1, \dots, \mathbf{X}_J$ , the design matrix  $\mathbf{C} = \{c_{j,k}\}$ , the function  $g$  and the shrinkage constants  $\tau_1, \dots, \tau_J$ ,

RGCCA optimizes the problem:

$$\max_{\mathbf{a}_1, \dots, \mathbf{a}_J} \sum_{j,k=1, j \neq k}^J c_{kj} g(\text{Cov}(\mathbf{X}_j \mathbf{a}_j, \mathbf{X}_k \mathbf{a}_k))$$

subject to the constraints  $\tau_j \|\mathbf{a}_j\|^2 + (1 - \tau_j) \text{Var}(\mathbf{X}_j \mathbf{a}_j) \quad j = 1, \dots, J$ , where the  $\mathbf{a}_j$  are the loading vectors associated to each block  $j$ .

# sGCCA

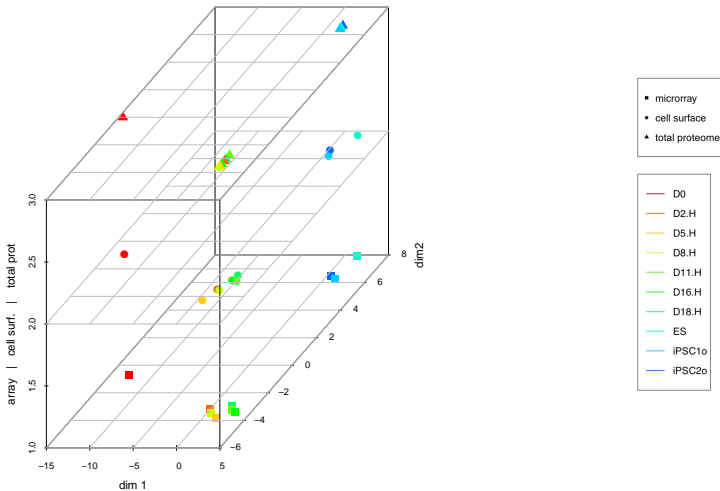
- Similar to the sPLS,  $L_1$  penalizations can be applied to the loading vectors to obtain a **sparse** version of RGCCA to select different types of biological entities across different functional levels

**Grandiose project:** Longitudinal study of cell reprogramming. In this study: 8 time points are considered. Multi platform study involving: 6 platforms: microarray, cell surface proteome, total proteome, RA-seq isoform, RNA-seq genes, miRNA.

**Tenehaus, A., et al.** [Variable Selection For Generalized Canonical Correlation Analysis](#), *submitted*.



## Integration of 3 levels



# Conclusions

- Statistical exploratory and integrative tools to extract patterns in time course data
- Can be applied to a variety of problems
- Does not provide p-values but can help generating new hypotheses, further statistical tests can then be applied
- Future directions: biological interpretation of the gene lists, time delay, generalised multi-way analysis, identifying discordant clusters across data sets for the same genes ...

## Neuronal time course

**Christine Wells** UQ

Ernst Wolvetang UQ

## Kidney transplant study

**Oliver Günther** UBC

Scott Tebutt UBC

## Grandiose proj. and Nagy group

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## Multiple data integration

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## Time course developments

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## FANTOM consortium

**sample providers** RIKEN, Japan

